

## Suitable Docking Protocol for the Design of Novel Coumarin Derivatives with Selective MAO-B Effects

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### Abstract

Recently, the application of molecular docking is drastically increasing due to the rapid growth of resolved crystallographic receptors with co-crystallized ligands. However, the inability of docking softwares to correctly score the occurred interactions between ligands and receptors is still a relevant issue. This study examined the Pearson's correlation coefficient between the experimental monoamine oxidase-B (MAO-B) inhibitory activity of 44 novel coumarins and the obtained GOLD 5.3 docking scores. Subsequently, optimization of the docking protocol was carried out to achieve the best possible pairwise correlation. Numerous modifications in the docking settings such as alteration in the scoring functions, size of the grid space, presence of active waters, and side-chain flexibility were conducted. Furthermore, ensemble docking simulations into two superimposed complexes were performed. The model was validated with a test set. A significant Pearson's correlation coefficient of 0.8217 was obtained for the latter. In the final stage of our work, we observed the major interactions between the top-scored ligands and the active site of 1S3B.

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## INTRODUCTION

MAO (monoamine oxidase) is an enzymatic system that comprises two isoenzymes: MAO-A and MAO-B, with more than 70% identity in their amino acid sequence<sup>1</sup>. The prominent role of MAO-A and MAO-B is the oxidation of aliphatic and aromatic amines to the corresponding aldehydes. The inhibition of the latter is implemented in two substantial neurodegenerative diseases – Alzheimer's and Parkinson's diseases<sup>2</sup>.

Administration of MAO-B inhibitors in treating the latter disorders has shown promising results in early and advanced stages of the conditions, considering the conservation of high dopamine levels in the synaptic cleft. Hence, both endogenous and exogenously administered dopamine concentrations could be retained after treatment with selective MAO-B inhibitors, and the effects are manifested<sup>3</sup>. Moreover,

all monoamine oxidase inhibitors demonstrate additional neuroprotective effects arising from decreased neuronal toxic radicals and peroxides<sup>4</sup>.

A significant breakthrough in the design and the optimization of novel MAO-B inhibitors has been made after the first resolved crystallographic MAO-B structure<sup>5</sup>. The study revealed three distinct domains in the active site of the receptor – entrance pocket, substrate cavity, and aromatic cage. It has also been postulated that the major ligand interactions in the binding gorge are the hydrophobic ones<sup>6</sup>. Moreover, four amino residues: Tyr-326, Leu-171, Ili-199, and Phe-168, are reported to act like a "gate" between the entrance and the substrate cavities. For an additional stabilization in the ligand-receptor complex, hydrogen bonds with Tyr-398<sup>7</sup>, Gln-206<sup>8</sup>, and FAD<sup>9</sup> have been described.

Ever since the computer-aided drug design (CADD) simulations were introduced in the drug discovery

processes, the time required to develop and optimize new molecules was drastically reduced<sup>10</sup>. Molecular docking is one of the most utilized structure-based drug design (SBDD) techniques as it is emerging as frequently applied in the optimization step of active ligands. Furthermore, it could be utilized for the virtual screenings of novel and effective drug candidates<sup>11,12</sup>. However, several challenges regarding the accuracy of molecular docking are still to be resolved. The major issues are related to the estimative character of the scoring functions<sup>13</sup> and the inefficiency of the fully flexible simulations<sup>14</sup>.

In order to acquire reliable docking results, the molecular docking protocol should be validated. Several validation methods such as re-docking, cross-docking, high enrichment factors, and enhanced correlation coefficients between experimental affinities and docking scores have been described<sup>15</sup>. The latest technique is often used to evaluate the correctness of the scoring function and the search algorithm through a created relationship with the experimentally acquired data. The correlation coefficients can vary considering the applied chemical dataset and the characteristics of the receptor<sup>16</sup>. However, an optimization of the docking protocol after altering the size of the binding pocket, the utilized scoring function, the flexibility of the side chain residues, and the presence of active waters often lead to superior correlation values<sup>17</sup>.

This study aimed to examine the effects of various docking protocols on Pearson's correlation coefficient of a coumarins dataset. All docking settings included in GOLD 5.3 were incorporated in the optimization process with further correlation coefficient calculations at each stage. Moreover, the significant intermolecular interactions between the top-scored ligands and the active site of 1S3B were examined.

## METHOD

### *Hardware and Software*

The docking simulations were carried out on an AMD Ryzen 5 3600 6-core 3.6GHz CPU, GeForce GTX 1060 3 GB GPU, 16 GB RAM installed memory, 64-bit Operating system on Windows 10 Pro. GOLD 5.3 (Genetic Optimization for Ligand Docking) from The Cambridge Crystallographic Data Centre (<https://www.ccdc.cam.ac.uk/solutions/csd-discovery/Components/gold/>)<sup>18</sup> was used for the

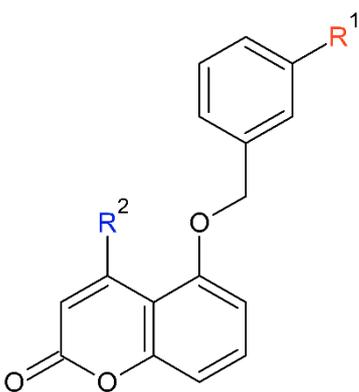
current docking simulations. It comprises four scoring algorithms: ChemPLP, GoldScore, ASP, and ChemScore. GoldScore considers mainly Van der Waals interactions and hydrogen bonds. ChemScore is the empirical algorithm of GOLD which was calibrated from numerous complexes with known binding affinities. ASP represents the knowledge-based function, while ChemPLP used piecewise linear potential to score the contacts in the ligand-receptor complex<sup>19</sup>.

In this study, all the latest scoring functions were utilized to evaluate the most prominent one for the current dataset. For the pre-docking procedures, the docking visualizer Hermes<sup>18</sup> was applied. The preparation of the ligands and the receptors were conducted in Hermes as well as ChemDraw and Chem3D from Perkin Elmer Informatics (<http://www.cambridgesoft.com/support/ProductHomePage.aspx?KBCatID=112>). The statistical calculations were performed in the JMP® Pro 12 from SAS Institute Inc. ([https://www.jmp.com/en\\_us/home.html](https://www.jmp.com/en_us/home.html)). The major interactions between the top-scored ligands and the active site of MAO-B receptor were visualized using Discovery Studio Visualizer from BIOVIA Dassault Systèmes, Pharmacopeia, Inc. (<https://www.3ds.com/products-services/biovia/products/molecular-modeling-simulation/biovia-discovery-studio/visualization/>).

### *Ligands*

Forty-four substituted coumarin derivatives were taken from a published paper by Pisani *et al*<sup>7</sup>. The ligands were grouped into two sets: a training and a test set. In the training set, we situated 35 compounds, while for the validation of the model, we applied nine structures. All coumarin derivatives used in our study were given in **Table I** with the corresponding pIC<sub>50</sub> values.

The drawing of the ligands and the conversion into the corresponding 3D structures were conducted in ChemDraw and Chem3D, respectively. The energy minimization procedures were also carried out in Chem3D with an early termination set at 2000 iterations and minimum root mean square (RMS) gradient fixed at 0.01000. During the docking simulations, the rotations of the ligands were set to "flexible".

**Table I.** Structures and pIC<sub>50</sub> values of the coumarin derivatives<sup>7</sup>


Training set			
Compound	R <sup>1</sup>	R <sup>2</sup>	pIC <sub>50</sub>
1	Cl	Et	7.54
2	Cl	Me	6.06
3	Cl	CH <sub>2</sub> Cl	7.33
4	Br	CH <sub>2</sub> Cl	6.38
5	Br	OH	6.31
6	Cl	CHO	7.28
7	Cl	COOEt	6.38
8	Cl	CONH <sub>2</sub>	6.63
9	Cl	CN	6.99
10	Cl	CH(OH)CH <sub>3</sub>	7.11
11	Cl	CH <sub>2</sub> CN	7.80
12	Cl	CH <sub>2</sub> CONHMe	7.62
13	Cl	CH <sub>2</sub> CON(Me) <sub>2</sub>	7.4
14	Cl	CH <sub>2</sub> NHMe	7.89
15	Cl	CH <sub>2</sub> N(Me) <sub>2</sub>	5.95
16	Cl	CH <sub>2</sub> -4'-morpholinyl	5.64
17	H	OMe	7.00
18	F	OMe	7.44
19	H	OEt	6.12
20	F	OEt	6.58
21	Cl	OEt	6.94
22	Br	OEt	6.9
23	Cl	OnPr	7.21
24	Br	OnPr	7.13
25	Cl	OCH <sub>2</sub> OMe	7.00
26	Cl	NHEt	7.55
27	Cl	NHCOOMe	7.41
28	Cl	NHCOOEt	6.23
29	Br	OCH <sub>2</sub> CONH <sub>2</sub>	80.08
30	Cl	OCH <sub>2</sub> CONHMe	7.47
31	Cl	(CH <sub>2</sub> ) <sub>2</sub> OH	8.13
32	Cl	(CH <sub>2</sub> ) <sub>2</sub> Cl	7.89
33	Cl	(CH <sub>2</sub> ) <sub>2</sub> Br	7.49
34	Cl	(CH <sub>2</sub> ) <sub>2</sub> CONH <sub>2</sub>	7.82
35	Cl	(CH <sub>2</sub> ) <sub>2</sub> CN	7.54
Test set			
1	Cl	OH	6.32
2	Cl	COCH <sub>3</sub>	7.40
3	Cl	CHNOH	6.66
4	Cl	CH <sub>2</sub> CONH <sub>2</sub>	7.52
5	H	OnPr	6.24
6	Cl	OCH <sub>2</sub> CON(Me) <sub>2</sub>	6.30
7	Cl	OCH <sub>2</sub> COCH <sub>3</sub>	7.57
8	Cl	NHCH <sub>2</sub> CONH <sub>2</sub>	7.31
9	Cl	COCH <sub>2</sub> Cl	6.4

### Receptors

Receptors with Protein Data Bank (PDB) IDs 1OJA, 1OJC<sup>20</sup>, 1S3B<sup>21</sup>, and 2V60<sup>22</sup> were retrieved from Protein Data Bank (<https://www.rcsb.org>) with resolutions under 2 Å<sup>23</sup>. Monomer B from all receptors was removed with the co-crystallized ligands and the co-factors lying in the corresponding monomer. In the case of present covalent bonds between the co-crystallized ligands and the co-factors, they were removed.

### Docking protocol

The GOLD wizard setup was utilized for rapid extraction of co-crystallized ligands and waters. Additional hydrogen bonds were added with the help of the former wizard. The search efficiency was set at 100% (default setting) with no early termination. During all dockings, the ligands were set to flexible, and initial energy minimization was carried out.

The starting docking protocol was built out of ChemPLP as a scoring function, 6 Å binding grid, no protein flexibility, and no active waters. All default parameters were altered to obtain a higher correlation coefficient between the experimental data and the obtained fitness scores. Primarily, the scoring functions and the size of the binding space were varied. The scoring algorithm was chosen based on the lowest R<sup>2</sup> value and the shortest simulation time. Analysis regarding the presence of active water molecules was performed, which examinations with and without waters in the active sites were conducted. After each simulation, the Pearson's correlation coefficient was calculated. After that, ten amino acids (Thr-195, Ile-198, Ile-199, Tyr-326, Phe-343, Leu-345, Tyr-398, Thr-399, Tyr-435, and Met-436) located in the binding cleft were set to flexible to examine the shift in the correlation coefficient. Finally, an ensemble docking was conducted after the superimposition of 1S3B-2V60 and 1OJA-1OJC-1S3B receptor structures. The latter complex demonstrated the highest MAO-B enrichment in one of our recent researches (to be published) and thus was examined in the current work.

## RESULTS AND DISCUSSION

### Re-docking

Re-docking procedures were carried out to assess the ability of the docking software to correctly place the co-crystallized ligands back into the binding pocket of the

receptors<sup>24</sup>. The reliability of GOLD 5.3 to correctly place the co-crystallized ligands of 1OJA, 1OJC, 1S3B, and 2V60 was unambiguous. The root-mean-square deviation (RMSD) for all receptors was under 2 Å, as shown in **Table II**.

**Table II.** RMSD values of the re-docked co-crystallized ligands of 1OJA, 1OJC, 1S3B and 2V60

PDB	Co-crystallized ligand	RMSD (Å)
1OJA	Isatin	1.18
1OJC	N-(2-Aminoethyl)-P-chlorobenzamide	1.25
1S3B	N-[(1S)-2,3-dihydro-1H-inden-1-yl]-N-methyl-N-prop-2-ynylamine	1.4
2V60	7-[(3-chlorobenzyl)oxy]-2-oxo-2H-chromene-4-carbaldehyde	1.1

### Optimizations of the docking protocol

All four scoring algorithms were applied to evaluate which was the most prominent one for the current dataset<sup>25</sup>. The rest of the docking settings were set to default. After each docking simulation, Pearson's correlation coefficient was calculated. ChemPLP had displayed the ability to acquire the highest correlation value, and the former scoring function was employed in the forthcoming protocols, as shown in **Table III**. Interestingly, GoldScore, ChemScore, and ASP showed significantly lowered accuracy when they were utilized to score the current dataset. GoldScore could not correctly score compound **16**, which led to a significantly lowered Pairwise correlation coefficient of 0.2802. Moreover, when ASP was used, there was no correlation at all –  $R^2=0.0876$ . The scoring function with the fastest run time was ChemPLP, while GoldScore demonstrated the longest simulation periods (double the time of ChemPLP). Considering the lengthy docking times, together with the unacceptable correlation coefficients obtained with GoldScore, ChemScore, and ASP, further studies with the latter GOLD 5.3 scoring algorithms were not conducted.

**Table III.** Calculated pairwise correlations of GOLD 5.3 scoring functions

Variable 1	Variable 2	Correlation	Count	Lower 95%	Upper 95%	Signif. Prob.
pIC <sub>50</sub>	ChemPLP 6 Å	0.5138	35	0.2178	0.7232	0.0016
pIC <sub>50</sub>	GoldScore 6 Å	0.2802	35	-0.0585	0.5611	0.1030
pIC <sub>50</sub>	Chemscore 6 Å	0.3535	35	0.0229	0.6143	0.0373
pIC <sub>50</sub>	ASP 6 Å	0.0876	35	-0.2531	0.4089	0.6169

Subsequently, the size of the binding gorge was modified to 8, 10, and 12 Å, when ChemPLP was used as a scoring function. Overall, when the size of the grid

box was expanded, the correlation coefficients increased<sup>26</sup>. The highest value ( $R^2=0.5929$ ) was obtained when the grid size was set to 12 Å. For the magnitude of the applied dataset, the run times were relatively similar, with a slight increase after each expansion of the grid space. The results were presented in **Table IV**.

**Table IV.** Calculated correlation coefficient after alterations in the size of the binding gorge

Scoring Algorithm	Grid size (Å)	Pearson's correlation coefficient
ChemPLP	6	0.5138
	8	0.5805
	10	0.5801
	12	0.5929

Optimization of the docking protocol was proceeded by altering the presence of water molecules in the active site<sup>27</sup>. Eight active water molecules (HOH-612, HOH-617, HOH-621, HOH-631, HOH-671, HOH-818, HOH-871, and HOH-883) were extracted and utilized for subsequent docking simulations. The calculated  $R^2$  value, when the waters were taken into consideration, was given in **Table V**. A drastic drop in the correlation coefficient was observed after the employment of active waters. In addition, the inability of the docking software to correctly score compound **34** was noticed. The former ligand received a false-positive fitness score of 121; thus, the correlation coefficient was calculated to be 0.4603. No further examinations with active waters were conducted considering the latter observations. It is important to note that in most cases, the active water molecules play an essential role in forming a stable complex<sup>28</sup>. However, in this work, the number of falsely scored results significantly increased when eight water molecules were included in the binding site.

**Table V.** Pairwise correlation coefficient after docking simulations with active waters

Variable 1	Variable 2	Correlation	Count	Lower 95%	Upper 95%	Signif. Prob.
pIC <sub>50</sub>	ChemPLP 12Å waters in the active site	0.4603	35	0.1501	0.6880	0.0054

During the last optimization step, we considered the flexibility of the side chain residues. Ten amino acids: Thr-195, Ile-198, Ile-199, Tyr-326, Phe-343, Leu-345, Tyr-398, Thr-399, Tyr-435, and Met-436, located in the active site, were set to a freely flexible state during the docking simulations. It was noted that the  $R^2$  value

dropped significantly from 0.5829 to 0.2921. The latter observation disposes a concern into the positive effect of flexible side chains in the reliable representations and scoring of the occurring MAO-B/coumarins intermolecular interactions. Furthermore, papers discussing higher enrichment values after semi-flexible receptor docking were reported<sup>29,30</sup>, which contrasted with this work.

In addition to the unsatisfactory results obtained from the side chain flexible dockings, the ensemble docking simulations of the described coumarins into two superimposed complexes: 1S3B-2V60 and 1OJA-1OJC-1S3B was examined. The receptor 2V60 was used owing to the chemical similarity between the co-crystallized ligand and the utilized in this study dataset. At the same time, the second superimposed complex demonstrated the highest enrichment value in a recently conducted study (to be published). Correlation coefficients of 0.4238 and 0.4508 were obtained, as shown in Table VI. However, the described technique was not applicable for a future virtual screening due to lower Pairwise correlation coefficients than the protocol mentioned earlier.

**Table VI.** Ensemble docking simulations into superimposed complexes

Variable 1	Variable 2	Correlation	Count	Lower 95%	Upper 95%	Signif. Prob.
pIC <sub>50</sub>	1S3B-2V60	0.4238	35	0.1054	0.6634	0.0112
pIC <sub>50</sub>	1OJA-1OJC-1S3B	0.4508	35	0.2134	0.6951	0.0093

Overall, the most prominent GOLD 5.3 docking protocol of novel MAO-B inhibitors with coumarin moiety was composed of the scoring function ChemPLP, size of the binding site 12 Å, absence of active waters, and no partial protein flexibility. Furthermore, the utilization of ensemble docking did not achieve any enhancements in the correlation value.

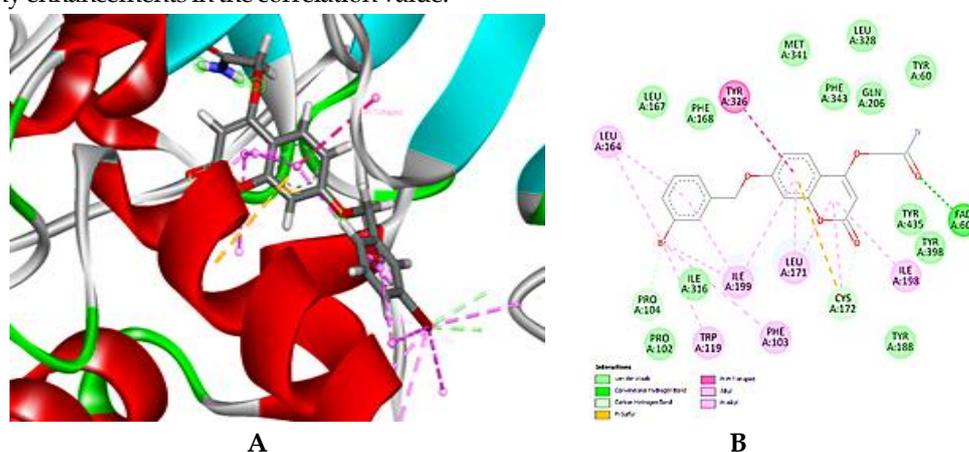
In order to validate the docking protocol, a test set built of nine chemically similar to the training set ligands, with a wide range of experimentally acquired binding affinities, was applied. The calculated Pearson's correlation coefficient of the test set equaled 0.8217, as shown in Table VII. The latter value was statically significant; thus, the model could be applied for a future virtual screening of novel MAO-B inhibitors.

**Table VII.** Ensemble docking simulations into superimposed complexes

Variable 1	Variable 2	Correlation	Count	Lower 95%	Upper 95%	Signif. Prob.
pIC <sub>50</sub>	Test set	0.8217	9	0.3470	0.9613	0.0066

### Visualizations of the major interactions

Two of the top-scored compounds located in the training set were visualized their major interactions with the active site of 1S3B. Both the 2D and 3D interaction diagrams of compounds **29** and **34** were provided in Figures 1 and 2, respectively. As demonstrated below, the poses of both ligands in the active site of MAO-B were exceptionally similar. The amide group in both cases was sandwiched in the aromatic cage, and a strong hydrogen bond was formed with FAD600. The core structure of coumarin was located in the substrate pocket, where it was stabilized by Van der Waals and hydrophobic interactions. The latter weak forces also occurred between the p-substituted phenyl moiety and the entrance cavity of the receptor. The absence of a delocalized cyclic system in the aromatic cage led to  $\pi$ - $\pi$  stacking interactions between the benzene ring<sup>31</sup> and the amino residue Tyr-326. A  $\pi$ -sulfur bond between Cys-172 and the coumarin's phenyl group in compound **29** was the only deviation in the interaction pattern between the two ligands.



**Figure 1.** (A) 2D and (B) 3D diagrams of the major occurring interactions between 1S3B and the compound **29**.

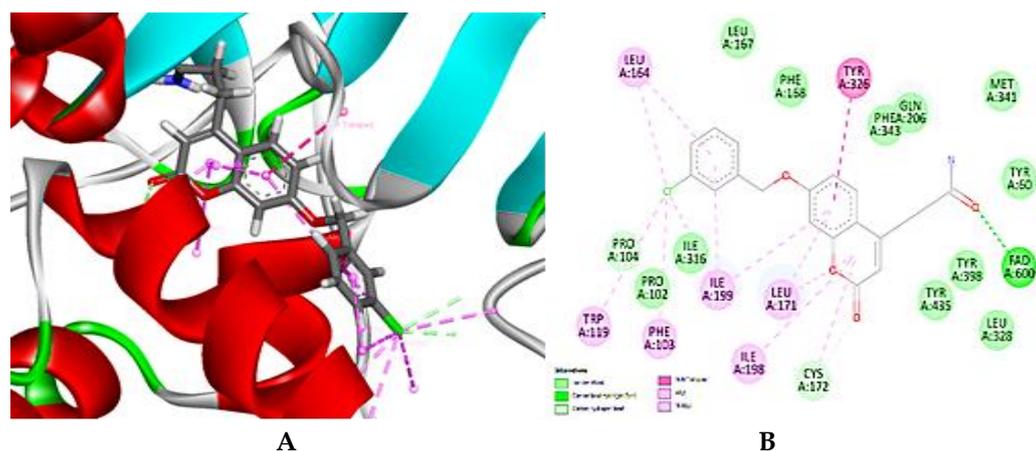


Figure 2. (A) 2D and (B) 3D diagrams of the major occurring interactions between 1S3B and the compound 34.

All the docking fitness scores and the major amino residues that participate in stabilizing the ligand-receptor complexes in the test set are given in **Table VIII**. As discussed before, nine ligands were included in the test set, and an  $R^2$  value of 0.8217 was achieved. Thus, all of the analyzed docking poses should be close to the actual poses of the ligands in the active site of MAO-B. Compound 4 displayed the highest fitness score of 92.11. The complex was stabilized with a hydrogen bond between FAD600 and the amide group. Moreover, a weaker carbon-hydrogen bond was present between Cys-172 and the pyran ring. Compound 1 showed the lowest score of 76.93, which was plausible considering the low number of stabilizing bonds.

Table VIII. Fitness scores and major interacting amino acid residues of the test set

Compound	Fitness score	Interacting amino acid residues
1	76.93	Cys-172, Ile-199, Ile-198, Leu-171, Leu-164, Trp-119
2	85.00	Tyr-326, Ile-199, Cys-172, Leu-171, Ile-198, Trp-119, Leu-164, Leu-167
3	80.07	FAD600, Tyr-326, Ile-199, Cys-172, Ile-198, Leu-171, Leu-164
4	92.11	FAD600, Cys-172, Tyr-326, Ile-199, Leu-171, Ile-198, Leu-167, Leu-164,
5	83.46	Tyr-326, Cys-172, Tyr-398, Ile-198, Ile-199, Leu-164, Ile-316
6	83.00	FAD600, Tyr-326, Ile-199, Trp-119, Leu-164, Leu-171, Ile-198
7	90.13	FAD600, Tyr-188, Phe-168, Cys-172, Tyr-326, Ile-199, Ile-198, Leu-164, Trp-119, Phe-103
8	86.29	Tyr-326, Cys-172, Pro-104, Ile-199, Ile-198, Leu-164, Phe-103, Leu-171, Leu-163
9	79.63	Tyr-326, Phe-168, Cys-172, Ile-199, Ile-198, Pro-104, Phe-103

## CONCLUSION

Good correlation coefficients were achieved in this work between the  $pIC_{50}$  values of 44 coumarins derivatives with MAO-B activity and their fitness scores applying the docking software GOLD 5.3. After optimizing the docking protocol for scoring functions, grid spaces, and rotatable side residues, a pairwise correlation of 0.5929 for the training set and 0.8217 for the test set was obtained. The presence of active waters and the inclusion of partial protein flexibility that was examined did not lead to enhanced correlation coefficients. In addition, compounds 29 and 34 demonstrated strongly identical poses in the active site of 1S3B. Overall, a statistically significant Pearson's correlation coefficient was obtained between coumarins derivatives and ChemPLP docking scores –  $R^2=0.5929$ . This finding could be beneficial for future virtual screenings in search of novel MAO-B inhibitors.

## CONFLICTS OF INTEREST

The authors have no conflicts of interest to declare that are relevant to the content of this article.

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None.

## DATA AVAILABILITY

All data are available from the authors.

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## AUTHORS' CONTRIBUTIONS

**Emilio Viktorov Mateev:** conceptualization, data curation, formal analysis, investigation, methodology, validation, visualization, writing – original draft, writing – review & editing. **Iva Valkova:** software, methodology, writing – original draft, writing – review & editing. **Maya Georgieva:** supervision, writing – original draft, writing – review & editing. **Alexander Zlatkov:** supervision, funding acquisition, writing – review & editing.

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